HOUSTON LEADING MEDICINE



Background

As a defining characteristic of a tumor, a cell's microenvironment plays an impose barriers for immune reaction by recruitment of other cells primarily by fibroblasts.[1] Fibroblast ability to manufacture collagen allows surrounding tumor tissue to stiffen by a more disorganized crosslinking of collagen fibers compared to normal tissue.[2] This adaptation to the cancer is what makes the fibroblast a primary factor to its development of cancer as well. Inactive or quiescent fibroblasts can be turned "cancer-associated fibroblasts" by unknown causes. Not only normal fibroblasts, but other mesenchymal cells are recruited.[4] These are the common aggressive behaviors that give cancer it's known dangerous connotation.

SCAFFOLD MAKING

Objectives

Develop a collagen solution that can provide an optimal environment for specified cancer cells. Pack the solution in a compact and uniform manner.

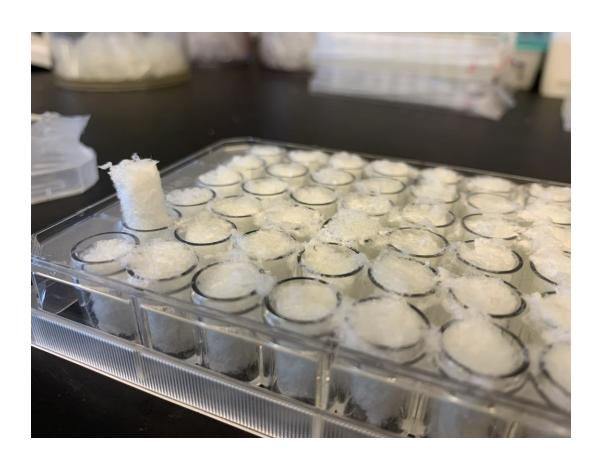
Methods

(1) Weigh out collagen and mix with milliQwater. (2) Adjust the pH of the new solution to approximately 5-6 pH using NaOH as a base to counter acids already in the collagen. (3) Sieve the solution three times using a 500micrometer sieve to rid it of preservatives. (4) Stir in BDDGE to strengthen the collagen by crosslinkage and leave for 24 hours in 4 degrees Celsius containment.

(5) Fill wells with developed collagen and freeze dry.

Results





Conclusions

- > Coculturing cancer cells with high ratio of stroma could increase their proliferation and metastatic potential
- > 3D models provide the newest technological insights in **preclinical drug evaluation**

Relationship Between Tumor Cells and Fibroblasts by Coculture of SKOV3 Ovarian Cancer and MRC5 Fibroblasts Jacob Wipf, Francesca Paradiso

SCAFFOLD **CHARACTERIZATION Objectives**

Prepare developed collagen scaffolds for scanning electron microscopy. Identify physical characteristics of models.

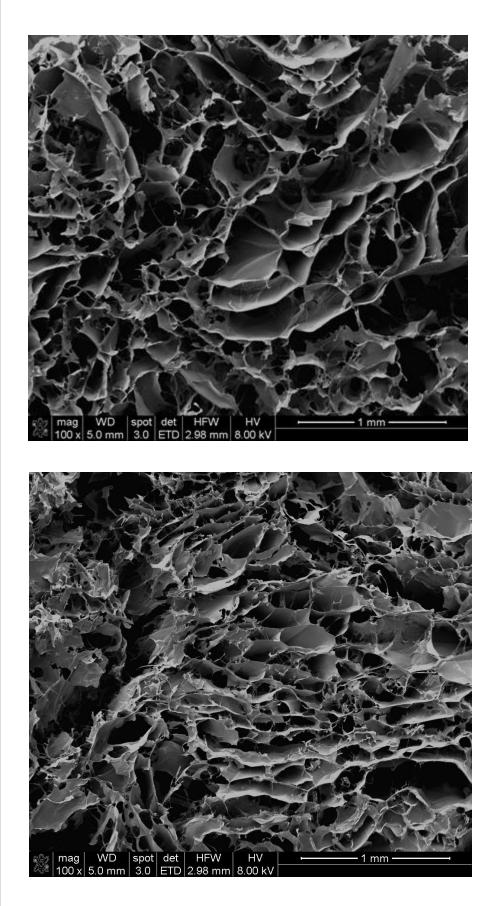
Methods

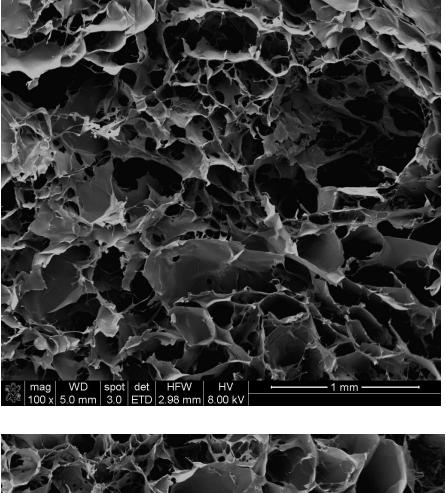
(1) Cover manufactured collagen scaffold with thin layer of platinum using vacuum sputter machine.

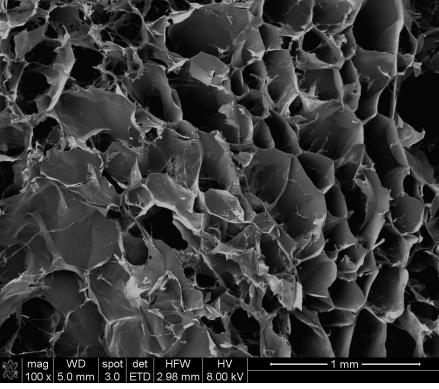
(2) View the scaffolds in a scanning electron microscope.

Results

SEM Images



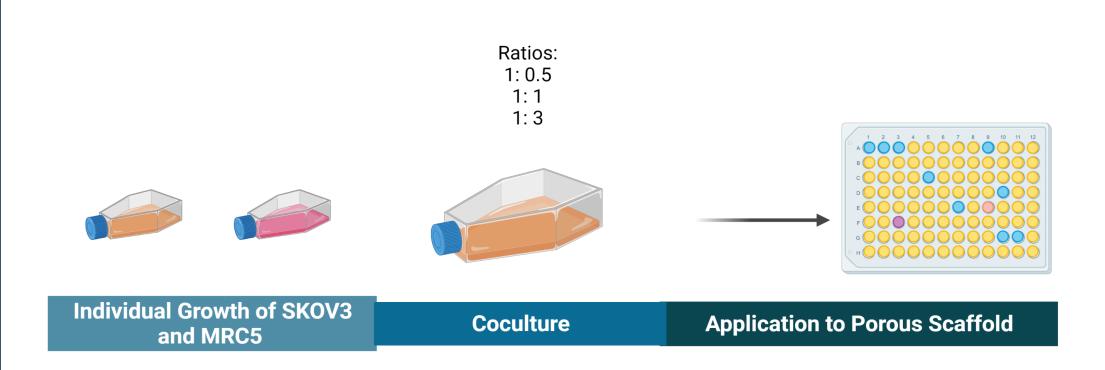






Provide space for cells to grow by periodically dividing the culture and providing more room to grow. Observe dyed cells on scaffold through immunofluorescence microscopy. Document cell growth among ratios.

Methods



> The 3D model used, a porous scaffold, has been an **informative tool** for examining developed **cocultures** of cancer and tumor stroma

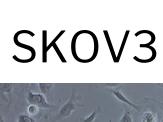


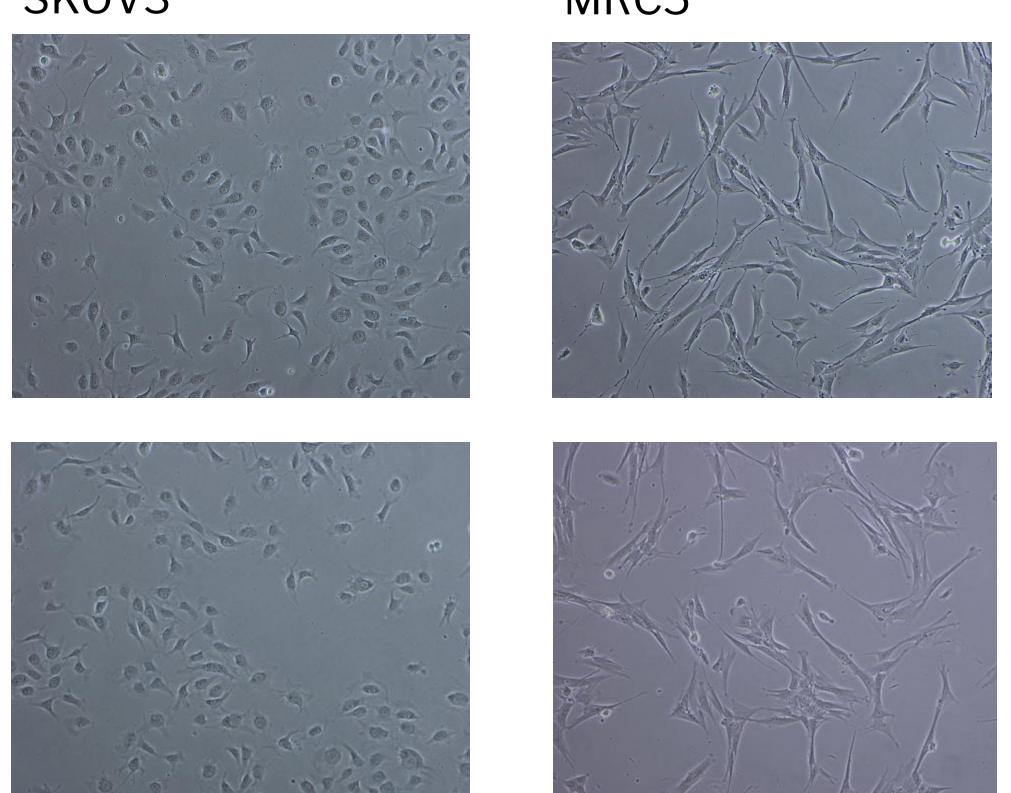
Cell culture & Coculture

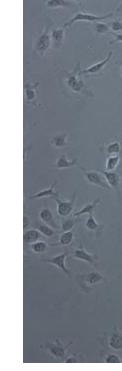
Objectives

- (1) Remove growth medium.
- (2) Add PBSH and trypsin. View cells
- detachment under microscope.
- (3) Insert into centrifuge.
- (4) Remove liquids to isolate cell pellet.
- (5) Dilute pellet with medium.
- (6) Add freezing agent and freeze overnight.



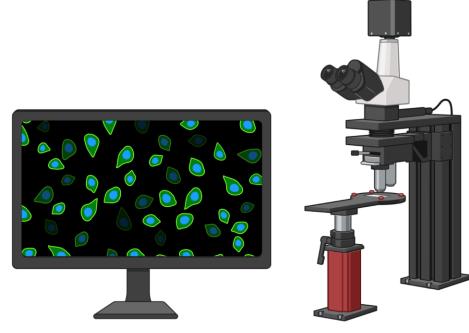






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(1) Apply and dye cells to scaffolds. (2) View under fluorescent microscope.



MRC5

References

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[2] Boyi Zhang, Fei Chen, Qixia Xu, Liu Han, Jiaqian Xu, Libin Gao, Xiaochen Sun, Yiwen Li, Yan Li, Min Qian, Yu Sun Revisiting ovarian cancer microenvironment: a friend or a

[3] Luke Bu, Hideo Baba, Naoya Yoshida, Keisuke Miyake, Tadahito Yasuda, Tomoyuki Uchihara, Patrick Tan & Takatsugu Ishimoto Biological heterogeneity and versatility of cancer-associated fibroblasts in the tumor microenvironment 2019

[4] João Rodrigues, Marcel A. Heinrich, Liliana Moreira Teixeira, and Jai Prakash <u>3D In</u> Vitro Model (R)evolution: Unveiling Tumor–Stroma Interactions 2020